

Study of the Effect of Plant Phytochemicals as Cdk-4 Inhibitor for the treatment of Retinoblastoma: An In-silico Approach

Mohana Priya I¹, Azar Zochedh A S^{2*}, Karthick A³

¹PG Student, Department of Bioinformatics, School of Distance Education, Bharathiar University, Coimbatore, Tamilnadu, India

²Research Scholar, Department of Biotechnology, Kalasalingam Academy of Research and Education, Krishnankoil, Tamilnadu, India

³M.Tech Biotechnology, Sole Proprietor, Madurai, Tamilnadu, India

Abstract - Retinoblastoma (RB) is the most common pediatric cancer of eye with an incidence of 1 in 15000 live births and accounts for 3% of all pediatric cancers. RB1, a tumor suppressor gene, encodes retinoblastoma protein (pRB) that regulates various cellular processes such as cell proliferation, differentiation, and apoptosis, and its inactivation results in uncontrollable cell proliferation. Mutations in RB1 gene leading to non-functional or loss of protein are reported in many human cancers including retinoblastoma. Due to involvement of Cyclin Dependent Kinases (Cdks) in the phosphorylation step, its inhibition leads to the activation of the suppressor activity of RB protein that prevents tumour progression. In this present study, we analyze the molecular docking studies of plant derived phytochemicals Naringin, Syringic acid and Berberine hydrochloride to find the binding efficiency of CDK-4 protein. The 3-dimensional structure of target protein is retrieved from the RCSB protein data bank and autodock vina tool helps in molecular docking studies. Docking analysis reveals that among three compounds Naringin shows good binding affinity of -7.7Kcal/mol with the active site of the CDK-4 protein and can be used as a potential drug to target retinoblastoma.

Whereas non-hereditary retinoblastoma, both hits occur in retinal progenitor cells only [4].

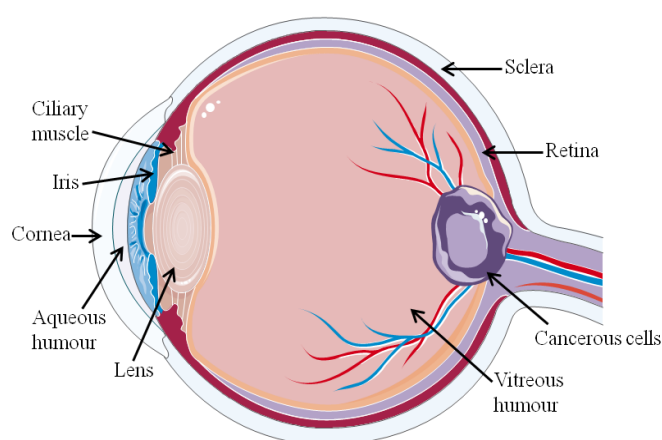


Fig -1: Anatomy of Retinoblastoma eye

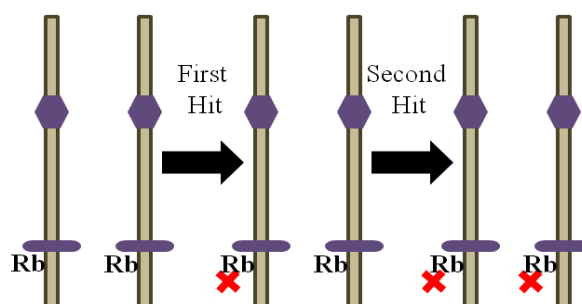


Fig -2: Sporadic RB - Two hit required

Key Words: Retinoblastoma, CDK-4, Naringin, Syringic Acid, Berberine Hydrochloride, Molecular docking

1. INTRODUCTION

1.1 Retinoblastoma

Retinoblastoma (RB) is the most common intraocular tumour arising from cone precursor cell with an incidence of 1 in 15000 live births [1]. It is estimated that India alone diagnoses 1500 new retinoblastoma cases every year [2]. Retinoblastoma is genetically characterized by mutation of both alleles of RB1 gene and inherited in an autosomal dominant pattern. Totally, 32 clinical signs were reported for the presence of RB. Leukocoria (white reflex) and strabismus are the most common presenting signs in more than 90% of RB cases. Other presentation signs include proptosis, red eye, orbital cellulitis, intraocular hemorrhage and reduction in vision. Retinoblastoma occurs as heritable or non-hereditary with one eye (unilateral) or both eyes (bilateral) affected [3]. In 1971, Knudson gave a hypothesis called two hit hypothesis that hereditary retinoblastoma, first hit is a germ line mutation and the second hit occurs in retinal progenitor cells (Fig -2).

1.2 RB1 Gene

RB1 gene is a tumour suppressor gene, located on the long arm of chromosome 13 in human. The retinoblastoma protein (pRB) regulates the cell cycle by targeting E2F transcription factor and function as transcriptional repressor. RB protein regulates various cellular processes such as cell proliferation, differentiation and apoptosis, and its inactivation results in uncontrollable cell proliferation [5]. Due to involvement of Cyclin Dependent Kinases (Cdks) in the phosphorylation step, its inhibition leads to the activation of the suppressor activity of RB protein that

prevents tumour progression. According to the previous study, it has been reported that CDK-4 plays a vital role in the phosphorylation of pRB and emerged as a planned focus to treat RB [6, 7].

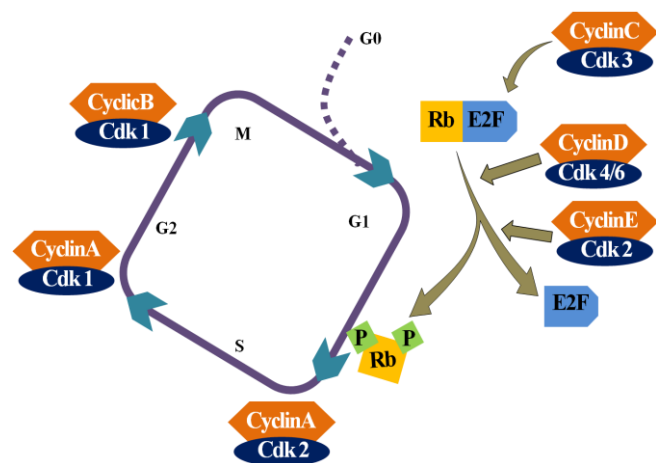


Fig -3: Function of Retinoblastoma protein

1.3 Naringin

Naringin is a flavanone glycoside compound present in grapes and citrus fruits and possesses a distinct bitter taste of grape juice and has strong antioxidant property [8]. The molecular weight of naringin is 80.4g/mol. The molecular and chemical formula are C₂₇H₃₂O₁₄ and 4',5,7-trihydroxyflavonone-7-rhamnoglucoside respectively [9,10]. Previous literature surveys have reported that naringin has a variety of pharmacological activity such as anti-cancer, anti-inflammation, anti-oxidation, anti-ulcer, anti-mutation, anti-osteoporotic and analgesic activities [11,12]. In addition, naringin can inhibit the excessive activation of the PI3K/AKT/mTOR signalling pathway which results in inhibiting biological function of tumors [13]. In recent survey, it was identified that PI3K/AKT/mTOR signalling pathway as a key target for tumor-targeted therapy [14-17]. These signalling pathway play a vital role in regulating proliferation, migration, growth and tumor cell survival [18,19]. Naringin is reported to show anticancer activity by enacting apoptotic pathway and promotes cell death in tumor cells to prevent the tumor growth with less adverse toxicity to normal cells [20].

1.4 Syringic Acid

Syringic acid is a naturally occurring phenolic compound distributed in wide variety of plant products (fruits and vegetables) as olives, spices, grapes, honey, pumpkin, red wine and certain fungal species [21-23]. The chemical formula and molar mass of Syringic acid are C₉H₁₀O₅ and 198.17 g/mol respectively. It is soluble in methanol or ethanol and is slightly soluble in water. The other names of Syringic acid are Cedar acid; 4-Hydroxy-3, 5-dimethoxybenzoic acid; Gallic acid; 3, 5-dimethyl ether; ; 3, 5-

Dimethoxy-4-hydroxybenzoic acid [24]. A previous report shows that, Syringic acid exhibits various properties in biomedical field such as anti-diabetic, anti-microbial, anti-cancer, anti-inflammation and also in prevention of brain/CNS, heart and liver [25]. Syringic acid also plays a major role in the industry sector, acts an effective substrate for fungal laccase enzyme, which has more importance in pulp industry and bioremediation [26]. Syringic acid derived from *Tamarix aucheriana* shows chemo sensitizing and anti-mitogenic activity in human colorectal cancer cells by involving in the mechanism of apoptosis regulation, cell-cycle arrest, NF κB- DNA binding, cell migration and proteasome activity [27]. Furthermore, literature reported that Syringic acid shows anti-proliferative activity against human K562 leukaemia cells [28].

1.5 Berberine Hydrochloride

Berberine hydrochloride is an isoquinoline alkaloid that has diverse pharmacological activity. It is derived from variety of Chinese herbs such as *Phellodendron chinense schneid*, *Phellodendron amurense* and *Coptidis rhizome*. It has anti-diabetic, anti-atherosclerotic action, anti-lipid peroxidation activity and also exhibits neuro-protective properties and enhance polycystic ovary syndrome [29-33]. It is widely used as an anti-fungal, anti-bacterial, and anti-inflammatory drug [34,35]. In various cancer cell lines berberine hydrochloride shows anti-proliferative activity that led to further research interest in this compound [36-45]. Berberine hydrochloride exhibits anti-neoplastic properties that induce apoptosis and cell cycle arrest and also inhibit cell migration and invasion through regulation of different pathways [46-49].

Computational biology strategies help to investigate the interaction between the therapeutic drugs and protein. This computer based study saves time and energy and also cost effective. The present study aims to characterize the physiochemical properties of target protein i.e., CDK-4 protein and to evaluate its binding affinity with naringin, syringic acid and berberine hydrochloride through molecular docking.

2. MATERIALS AND METHODS

2.1 Preparation of the 3-dimensional structure of protein

The 3D model of protein CDK-4 was retrieved from RCSB Protein Data Bank (PDB) (<https://www.rcsb.org/>), (PDB ID: 2w9z) was utilized as a potential drug target for retinoblastoma. The molecule structure of target protein was retrieved in “.pdb” format.

2.2 Ligand identification

Naringin, Syringic acid and Berberine hydrochloride (PubChem ID: 442428, 10742 and 12456) were compounds used for molecular docking study and were obtained from phytochemicals of various plants. These ligand molecules

were retrieved in 3-dimensional structure in “.sdf” format from PubChem (<https://pubchem.ncbi.nlm.nih.gov/>).

2.3 Physiochemical characterization of the protein model

The protein sequence in FASTA format was obtained from UniProt (UniProt ID: P24385) (<http://www.uniprot.org>). The obtained FASTA sequence was uploaded in ExPasy's ProtParam Proteomics server to analyze physical and chemical parameters of the target protein (<http://web.expasy.org/protparam>). The molecular weight, amino acid length, pI, extinction coefficient, positive and negative charged residues, grand average of hydropathicity index (GRAVY), instability and aliphatic index of the target protein has been identified.

2.4 Molecular docking through AutoDock vina

The molecular docking was performed using freely available software AutoDock Vina. The target protein was loaded in “.pdb” format and was prepared by adding hydrogen polar atoms and kollman charges and deleting water molecules. The prepared protein was finally saved in “.pdbqt” format. After preparing target protein the ligand molecules were imported in “.sdf” format and were converted to “.pdbqt” format. Then docking region was selected by forming a grid box. After that AutoDock Vina was executed using command prompt and results were analyzed

2.5 Construction of protein-ligand complex

The complex structure of target protein and ligand was built using PyMol 2.4 tool (<https://pymol.org/2/>). Both the target protein CDK-4 and ligand molecule were imported from the docking workspace. The imported file was in the “.pdbqt” format. Then the construction of protein-ligand complex was made and was saved in “.pdb” format.

2.6 Structure visualization

The protein-ligand complex was visualized using BIOVIA Discovery studio tool 2020. The constructed protein-ligand complex was imported on the graphical window in “.pdb” format. After loading the complex structure charges were added. The complex molecule showed the interaction of amino acid between target protein and ligand in 2-dimension and 3-dimension. Through 2D and 3D structure the hydrogen and hydrophobic bonds were analyzed.

3. RESULTS AND DISCUSSION

3.1 Physiochemical properties of target protein

The ExPasy's ProtParam tool was used to analyze the physiochemical characteristics of target protein CDK-4. The result is given in table 1.

Table -1: Physiochemical properties of target protein

Target Protein: CDK-4 (PDB ID: 2w9z)	
Length	295
Molecular weight	33729.11
pI	4.97

-R	47
+R	34
Extinction co-efficient at 280nm	20690
Instability index	57.71
Aliphatic index	92.92
GRAVY	-0.185

The isoelectric point (pI) value calculated for the target protein was less than 7 that shows the acidic characteristic of CDK-4 protein. The molecular weight of protein was 33729.11 Da. The extinction coefficient was calculated at 280nm using the extent of light being absorbed by the protein of target at wavelength range of 20690 M⁻¹cm⁻¹. -R and +R denotes the negative (ASP+GLU) and positive (ARG+LYS) charged residues. There are 47negative charged residues and 34 positive charged residues. Based on the ExPasy's ProtParam instability index of CDK-4 was 57.71. The grand average hydropathicity (GRAVY) index was negative value of -0.185 demonstrate the hydrophilic nature of the target protein.

3.2 Docking scores of target protein with plant phytochemicals

Further the docking was performed for protein CDK-4 against the plant derived phytochemicals naringin, syringic acid and berberine hydrochloride.

Table -2: AutoDock Vina results of Naringin, Syringic acid and Berberine hydrochloride against CDK-4

Ligand Molecules	Binding Affinity (kcal/mol)
Naringin	-7.7
Syringic acid	-5.9
Berberine hydrochloride	-6.9

The protein of target CDK-4 was docked on the binding pocket with the plant derived phytochemicals naringin, syringic acid and berberine hydrochloride. Based on the lowest energy value (DGbind) and negative value, the best docking orientation was selected. According to the AutoDock Vina result it was analyzed that syringic acid (-5.9kcal/mol) and berberine hydrochloride (-6.9kcal/mol) showed less binding affinity compared to naringin. Naringin shows strong interaction at the active sites and best docking confirmation towards the CDK-4 protein with docking score of -7.7 kcal/mol.

3.3 Analysis of 2D and 3D interaction of target protein with ligands

Our target protein CDK-4 with potent drug naringin formed four (4) Conventional Hydrogen bond interactions with Arg26 (2) and His68 (2) at bond distances 2.11196 Å, 2.77127 Å, 2.47646 Å, 2.20607 Å and one (1) Carbon Hydrogen bond with Phe130 at distance 3.68969 Å.

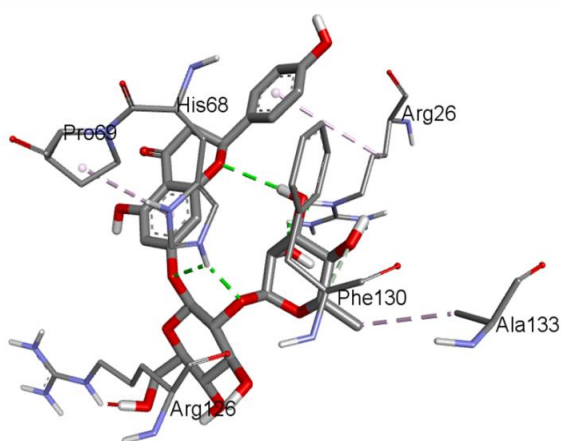


Fig -4: 3D Interaction of Naringin with CDK-4

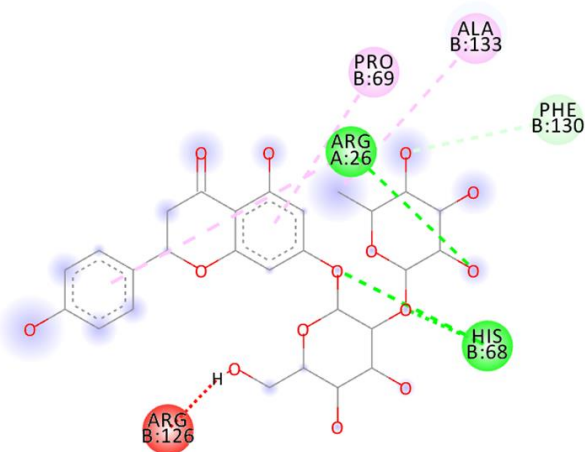


Fig -5: 2D Interaction of Naringin with CDK-4

Also CDK-4 with naringin, whereas alkyl type of hydrophobic interaction was formed at Ala133 with a distance 4.07757 Å and also formed two (2) Pi-Alkyl interactions with Pro69 and Arg26 at bond distances 5.12332 Å and 4.87918 Å respectively. Other interactions include unfavourable donar-donar interaction with Arg126 at distance 2.80802 Å. Thus, the compound naringin was found to be deeply buried in the active site of the ATP binding domain.

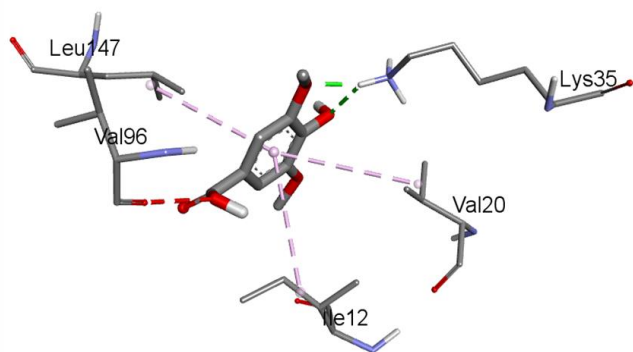


Fig -6: 3D Interaction of Syringic acid with CDK-4

Syringic acid with target protein CDK-4 formed two (2) Conventional Hydrogen bond interactions with Lys35 (2) at distance 1.96122 Å and 2.37577 Å respectively. Also syringic acid with target protein formed three (3) hydrophobic such as Pi-Alkyl interactions with amino acid residues of Ile12, Val20 and Leu147 at 5.03755 Å, 5.14824 Å and 5.04707 Å bond distances. In addition, syringic acid with CDK-4 formed unfavourable acceptor-acceptor interaction with Val96 amino acid residue.

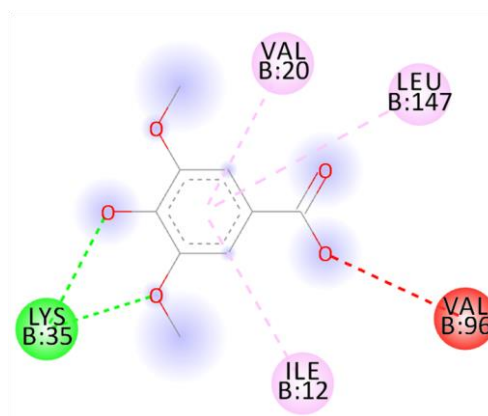


Fig -7: 2D Interaction of Syringic acid with CDK-4

Berberine hydrochloride with target protein formed one (1) Conventional Hydrogen bond interaction with amino acid residue Arg163 at bond distance of 3.05627 Å and one (1) Carbon Hydrogen bond interaction with Glu56 at distance 3.74346 Å.

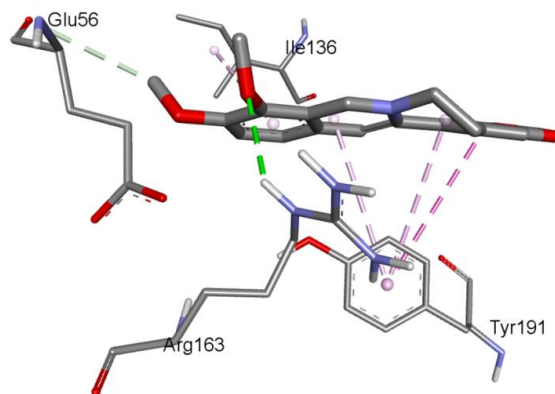


Fig -8: 3D Interaction of Berberine hydrochloride with CDK-4

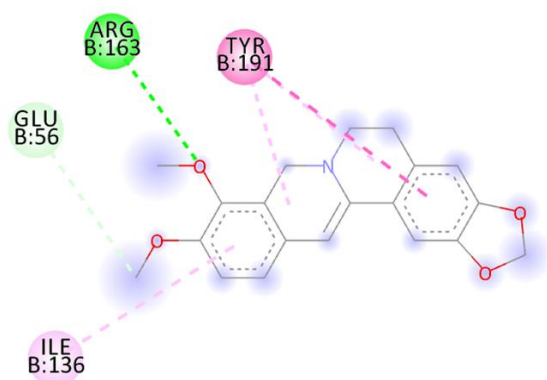


Fig -9: 2D Interaction of Berberine hydrochloride with CDK-4

Whereas, the amino residues Tyr191 (3) and Ile 136 formed one (1) Pi-Pi type and three (3) Pi-Alkyl type of hydrophobic interaction at 5.67776 Å, 5.29779 Å, 5.25385 Å and 5.34719 Å respectively.

Most of the docking studies have reported that binding affinity, number hydrogen bond interaction and bond distances play a major role on influencing protein-ligand interaction [50]. It shows that all the three ligand molecules exhibit good affinity towards the ATP binding site of CDK-4 protein. This could be easily interpreted based on the binding affinity with the CDK-4 protein makes naringin more potent drug than syringic acid and berberine hydrochloride.

4. CONCLUSION

In this study, with the help of molecular docking we have effectively elucidated the effect of plant derived phytochemicals naringin, syringic acid and berberine hydrochloride on the ATP binding site of CDK-4 protein. Among three ligands naringin shows higher binding affinity with CDK-4 with docking score of -7.7kcal/mol. The study can be useful to discover novel inhibitors and to design and develop drugs by validating in-vitro and in-vivo targeting retinoblastoma protein for the treatment of retinoblastoma.

REFERENCES

- [1] Dimaras H, Corson TW, Cobrinik D, White A, Zhao J, Munier FL, et al. Retinoblastoma. Nature reviews Disease primers. 2015.1:15021.
- [2] Gaikwad N, Vanniarajan A, Husain A, Jeyaram I, Thirumalairaj K, Santhi R, et al. Knudson's hypothesis revisited in Indian retinoblastoma patients. Asia-Pacific journal of clinical oncology. 2015.11:299-307.
- [3] Chawla B, Hasan F, Azad R, Seth R, Upadhyay AD, Pathy S, et al. Clinical presentation and survival of retinoblastoma in Indian children. British Journal of Ophthalmology. 2016.100:172-178.
- [4] Knudson AG, Jr. Mutation and cancer: statistical study of retinoblastoma. Proceedings of the National Academy of Sciences of the United States of America. 1971.68:820-823.
- [5] Goodrich DW. The retinoblastoma tumour-suppressor gene, the exception that proves the rule. Oncogene. 2006.25:5233-5243.

- [6] Spring L, Bardia A, Modi S. Targeting the Cyclin D/Cyclin-Dependent Kinase (CDK) 4/6-retinoblastoma pathway with selective CDK 4/6 inhibitors in hormone receptor-positive breast cancer: rationale, current status, and future directions. Discov Med. 2016.21:65-74.
- [7] Yu B, Lane ME, Pestell RG, Albanese C, Wadler S. Downregulation of cyclin D1 alters cdk 4- and cdk 2-specific phosphorylation of retinoblastoma protein. Mol cell Biol Res Commun. 2000.3: 352-359.
- [8] Chen R, Qi QL, Wang MT and Li QY: Therapeutic potential of naringin: An overview. Pharm Biol. 2016. 54: 3203-3210.
- [9] Azar Zochedh A S, Asath Bahadur S, Thandavarayan Kathiresan. Quantum chemical and Molecular docking studies of Naringin: A potent anti-cancer drug. Journal of Cardiovascular Disease Research. 2021.12:1140-1148.
- [10] Hongyun Cheng, Xue Jiang, Qian Zhang, Jun Ma, Ronghui Cheng, Hongmei Yong, Huichang Shi1, Xueyi Zhou, Liyue Ge, Guangyi Gao. Naringin inhibits colorectal cancer cell growth by repressing the PI3K/AKT/mTOR signaling pathway. Experimental and Therapeutic Medicine. 2020.19: 3798-3804.
- [11] Zhu H, Gao J, Wang L, Qian K, Cai L. In vitro study on reversal of ovarian cancer cell resistance to cisplatin by naringin via the nuclear factor- κ B signaling pathway. Exp Ther Med.2018.15:2643-2648.
- [12] El-Desoky AH, Abdel-Rahman RF, Ahmed OK, El-Beltagi HS, Hattori M. 2018. Anti-inflammatory and antioxidant activities of naringin isolated from Carissa carandas L.: in vitro and in vivo evidence. Phytomedicine. 2018.15:126-134.
- [13] Raha S, Yumnam S, Hong GE, Lee HJ, Saralamma VV, Park HS, Heo JD, Lee SJ, Kim EH, Kim JA, et al: Naringin induces autophagy-mediated growth inhibition by downregulating the PI3K/Akt/mTOR cascade via activation of MAPK pathways in AGS cancer cells. Int J Oncol. 2015. 47:1061-1069.
- [14] Morgensztern D and McLeod HL: PI3K/Akt/mTOR pathway as a target for cancer therapy. Anticancer Drugs. 2005. 16:797-803.
- [15] Scartozzi M, Giampieri R, Maccaroni E, Mandolesi A, Biagetti S, Alfonsi S, Giustini L, Loretelli C, Faloppi L, Bittoni A, et al: Phosphorylated AKT and MAPK expression in primary tumours and in corresponding metastases and clinical outcome in colorectal cancer patients receiving irinotecan-cetuximab. J Transl Med. 2012.10:71.
- [16] Oikonomou E and Pintzas A: Cancer genetics of sporadic colorectal cancer: BRAF and PI3KCA mutations, their impact on signaling and novel targeted therapies. Anticancer Res. 2006. 26A:1077-1084.
- [17] Francipane MG and Lagasse E: mTOR pathway in colorectal cancer: An update. Oncotarget. 2014. 5:49-66.
- [18] Kang S, Dong SM, Kim BR, Park MS, Trink B, Byun HJ and Rho SB: Thioridazine induces apoptosis by targeting the PI3K/Akt/mTOR pathway in cervical and endometrial cancer cells. Apoptosis. 2012. 17:989-997.
- [19] Hamada K, Sasaki T, Koni PA, Natsui M, Kishimoto H, Sasaki J, Yajima N, Horie Y, Hasegawa G, Naito M, et al: The PTEN/PI3K pathway governs normal vascular development and tumor angiogenesis. Genes Dev. 2005. 19:2054-2065.

- [20] Zeng L, Zhen Y, Chen Y, Zou L, Zhang Y, Hu F, Feng J, Shen J and Wei B: Naringin inhibits growth and induces apoptosis by a mechanism dependent on reduced activation of NF- κ B/COX-2-caspase-1 pathway in HeLa cervical cancer cells. *Int J Oncol.* 2014. 45:1929-1936.
- [21] J.M. Pezzuto, Grapes and human health: a perspective. *J Agric Food Chem.* 2008. 56:6777-6784.
- [22] A.M.G. Paramas, J.A. Gomez Bareza, C. Cordon Marcosa, J. Rafael García Villanova, J. Sánchez, Sánchezb, HPLC-fluorimetric method for analysis of amino acids in products of the hive (honey and bee-pollen). *Food Chemistry.* 2006. 95:148-156.
- [23] N. Sakaguchi, M. Inoue, Y. Ogihara, Reactive oxygen species and intracellular Ca²⁺, common signals for apoptosis induced by gallic acid. *Biochemical Pharmacology.* 1998. 55:1973-1981.
- [24] Srinivasulu Cheemanapallia, Ramgopal Mopurib, Ramanjaneyulu Gollac, Anuradha C.M.b, Suresh Kumar Chittaa. Syringic acid (SA) – A Review of Its Occurrence, Biosynthesis, Pharmacological and Industrial Importance. *Biomedicine & Pharmacotherapy.* 2018. 108:547-557.
- [25] P. Kiran, M. Denni, M. Daniel. Antidiabetic principles, phospholipids and fixed oil of kodo millet (*Paspalum scrobiculatum* Linn.). *Ind J Appl Res.* 2014. 4:13-15.
- [26] T. Abe, E. Masai, K. Miyauchi, Y. Katayama, M. Fukuda, Tetrahydrofolate Dependent O-Demethylase, LigM, Is Crucial for Catabolism of Vanillate and Syringate in *Sphingomonas paucimobilis* SYK-6. *J of Bacteriology.* 2005. 2030-2037.
- [27] M.S. Abaza, R. Al-Attayah, R. Bhardwaj, G. Abbadi, M. Koyippally, M. Afzal. Syringic acid from *Tamarix aucheriana* possesses antimutagenic and chemosensitizing activities in human colorectal cancer cells. *Pharm Biol.* 2013. 51:1110-1124.
- [28] S. Cheemanapalli, C.M. Anuradha, P. Madhusudhana, M. Mahesh, P.B. Raghavendra, C.S. Kumar, Exploring the binding affinity of novel syringic acid analogues and critical determinants of selectivity as potent proteasome inhibitors, *Anticancer Agents Med Chemistry.* 2016. 16:1496-1510.
- [29] Lee IA, Hyun YJ, Kim DH. Berberine ameliorates TNBS-induced colitis by inhibiting lipid peroxidation, enterobacterial growth and NF- κ B activation. *Eur J Pharmacol.* 2010. 648:162-170.
- [30] Liu X, Li G, Zhu H, et al. Beneficial effect of berberine on hepatic insulin resistance in diabetic hamsters possibly involves in SREBPs, LXR α and PPAR α transcriptional programs. *Endocr J.* 2010. 57:881-893.
- [31] Zhou J, Zhou S. Berberine regulates peroxisome proliferator-activated receptors and positive transcription elongation factor b expression in diabetic adipocytes. *Eur J Pharmacol.* 2010. 649:390-397.
- [32] Wu M, Wang J, Liu LT. Advance of studies on anti-atherosclerosis mechanism of berberine. *Chin J Integr Med.* 2010.16:188-192.
- [33] Zhao L, Li W, Han F, et al. Berberine reduces insulin resistance induced by dexamethasone in theca cells in vitro. *Fertil Steril.* 2011. 95: 461-463.
- [34] Remppis A, Bea F, Greten HJ, et al. Rhizoma coptidis inhibits LPS-induced MCP-1/CCL2 production in murine macrophages via an AP-1 and NF κ B-dependent pathway. *Mediators Inflamm.* 2010.194896.
- [35] Liu F, Liang HL, Xu KH, Tong LL, Tang B. Supramolecular interaction of ethylenediamine linked beta-cyclodextrin dimer and berberine hydrochloride by spectrofluorimetry and its analytical application. *Talanta.* 2007.74:140-145.
- [36] Choi MS, Yuk DY, Oh JH, et al. Berberine inhibits human neuroblastoma cell growth through induction of p53-dependent apoptosis. *Anticancer Res.* 2008.28:3777-3784.
- [37] Ho YT, Lu CC, Yang JS, et al. Berberine induced apoptosis via promoting the expression of caspase-8, -9 and -3, apoptosis-inducing factor and endonuclease G in SCC-4 human tongue squamous carcinoma cancer cells. *Anticancer Res.* 2009.29:4063-4070.
- [38] Hsu WH, Hsieh YS, Kuo HC, et al. Berberine induces apoptosis in SW620 human colonic carcinoma cells through generation of reactive oxygen species and activation of JNK/p38 MAPK and FasL. *Arch Toxicol.* 2007.81:719-728.
- [39] Patil JB, Kim J, Jayaprakasha GK. Berberine induces apoptosis in breast cancer cells (MCF-7) through mitochondrial-dependent pathway. *Eur J Pharmacol.* 2010.645:70-78.
- [40] Auyeung KK, Ko JK. *Coptis chinensis* inhibits hepatocellular carcinoma cell growth through nonsteroidal anti-inflammatory drug-activated gene activation. *Int J Mol Med.* 2009.24:571-577.
- [41] Yu FS, Yang JS, Lin HJ, et al. Berberine inhibits WEHI-3 leukemia cells in vivo. *In Vivo.* 2007;21(2):407-412.
- [42] James MA, Fu H, Liu Y, Chen DR, You M. Dietary administration of berberine or Phellodendron amurense extract inhibits cell cycle progression and lung tumorigenesis. *Mol Carcinog.* 2011.50:1-7.
- [43] Kim DW, Ahan SH, Kim TY. Enhancement of arsenic trioxide (As(2) O(3))-mediated apoptosis using berberine in human neuroblastoma SH-SY5Y cells. *J Korean Neurosurg Soc.* 2007.42:392-399.
- [44] Shen N, Li CN, Huan Y, Shen ZF. [Advances of the mechanism study on berberine in the control of blood glucose and lipid as well as metabolism disorders.] *Yao Xue Xue Bao.* 2010.45:699-704.
- [45] Zhang Q, Xiao X, Feng K, et al. Berberine moderates glucose and lipid metabolism through multipathway mechanism. *Evid Based Complement Alternat Med.* 2010. [Epub ahead of print].
- [46] Singh T, Vaid M, Katiyar N, Sharma S, Katiyar SK. Berberine, an isoquinoline alkaloid, inhibits melanoma cancer cell migration by reducing the expressions of cyclooxygenase-2, prostaglandin E and prostaglandin E receptors. *Carcinogenesis.* 2011.32:86-92.
- [47] Li-Weber M. Targeting apoptosis pathways in cancer by Chinese medicine. *Cancer Lett.* 2010. [Epub ahead of print].
- [48] Tsang CM, Lau EP, Di K, et al. Berberine inhibits Rho GTPases and cell migration at low doses but induces G2 arrest and apoptosis at high doses in human cancer cells. *Int J Mol Med.* 2009.24:131-138.
- [49] Zhang X, Gu L, Li J, et al. Degradation of MDM2 by the interaction between berberine and DAXX leads to potent apoptosis in MDM2- overexpressing cancer cells. *Cancer Res.* 2010.70:9895-9904.
- [50] Fuks, F., Burgers, W. A., Brehm, A., Hughes-Davies, L., & Kouzarides, T. DNA methyltransferase Dnmt1 associates with histone deacetylase activity. *Nature genetics.* 2000. 24:88-91.